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Comoclathris typhicola, a new species for the funga of Iran

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The genus *Comoclathris* Clem., typified by *Comoclathris lanata* Clem., was described by Clements (1909). It was placed in the family Pleosporaceae (Pleosporales, Dothideomycetes, Pezizomycotina, Ascomycota) based on *Alternaria*-like asexual morphs (Zhang et al. 2011, Woudenberg et al. 2013). Molecular data is not available for the type species, *C. lanata*, but the two putative strains of *C. compressa* (CBS 157.53 and CBS 156.53) cluster together in a well-supported clade within the family Pleosporaceae (Ariyawansa et al. 2014). Subsequent molecular studies confirmed the placement of *Comoclathris* in Pleosporaceae (Ariyawansa et al. 2015, Wijayawardene et al. 2017, Boonmee et al. 2021, Crous et al. 2021, Mattoo et al. 2023). The genus is characterized by ascomata with circular lid-like openings and applanate reddish-brown to dark reddish-brown, muriform ascospores, with single longitudinal septa (Ariyawansa et al. 2015, Wijayawardene et al. 2017). Presently, the genus consists of 40 registered names in Index Fungorum (https://www.indexfungorum.org; accessed Oct 2023).

In this study, leaf samples from *Typha latifolia* L. (Typhaceae, Poales) showing necrotic lesions were collected from Miyandoab city, West Azarbaijan province, Iran, in 2021. Leaf samples were surface disinfected using 1% sodium hypochlorite solution for three min, followed by rinsing in sterile distilled water and incubated in a moist chamber at 25 °C. The incubated leaves were inspected under the stereo microscope (SZ51, Olympus, Japan) and single-spore isolation was done following the method described in Ahmadpour et al. (2021). Germinated spores were transferred to potato dextrose agar (PDA: 39 g/L sterile distilled water, Merck, Darmstadt, Germany) plates and incubated at room temperature for 2–4 weeks. The isolates were grown on PDA, malt extract agar (MEA) and oatmeal agar (OA) culture media at 25 °C under the near ultraviolet light (NUV)/dark cycle of 12/12 h for 7–14 days to study the morphological characteristics (Mattoo et al. 2023). Measurements and microphotographs were prepared from slide mounts in lactophenol using an Olympus AX70 compound microscope with differential interference contrast (DIC) illumination. Adobe Photoshop 2020 v. 2.10.8 software (Adobe Inc., San Jose, California) was used for manual editing. All the identified isolates were deposited in the fungal culture collections of the Iranian Research Institute of Plant Protection (IRAN) and Urmia University (FCCUU). DNA was extracted from the mycelial mass of each isolate harvested from 10-days-old PDA Petri dishes using chloroform extraction and isopropanol precipitation method (Ahmadpour et al. 2021). Amplification and sequencing of parts of *RPB2* gene was carried out using RPB2-5F2/RPB2-7cr2 primer pairs. Maximum likelihood (ML) analysis was conducted in the RAxML–HPC BlackBox v. 8.2.8 (Stamatakis 2014) online server of the CIPRES Science gateway portal (https://www.phylo.org/) (Miller et al. 2012) for 1000 bootstrapping iterations, using the general time reversible model (GTR) with a discrete gamma distribution. Sequences of *Neocamarosporium chenopodii* (CBS 206.80) and *N. obiones* (CBS 432.77) served as the outgroup taxa (Mattoo et al. 2023). The resultant phylogenetic trees were visualized in FigTree v. 1.4.4 (Rambaut 2019), and edited in graphic design software, Adobe Illustrator® CC 2020. The newly generated sequences were submitted to GenBank. The resulting phylogram (Fig. 1) revealed that our isolates clustered well with *Comoclathris typhicola* (CBS 132.69 strain) in a distinct clade with high bootstrap support values (ML = 100). To the best of our knowledge, this is the first report of *C. typhicola* for the funga of Iran.

Comoclathris typhicola (Cooke) Ariyaw. & K.D. Hyde, Fungal Diversity 71: 105 (2015) **Fig. 2**

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Basionym: *Sphaeria typhicola* Cooke [as 'typhaecola'], Grevillea 5(no. 35): 121 (1877).
Synonyms: *Clathrospora typhicola* (Cooke) Höhn., Annls mycol. 16(1/2): 88 (1918), *Macrospora typhicola* (Cooke) Shoemaker & C.E. Babc., Can. J. Bot. 70(8): 1644 (1992), *Pleospora typhicola* (Cooke) Sacc., Reliq. Libert 2: no. 152 (1881), *Pyrenophora typhicola* (Cooke) E. Müll., Sydowia 5(3-6): 256 (1951).

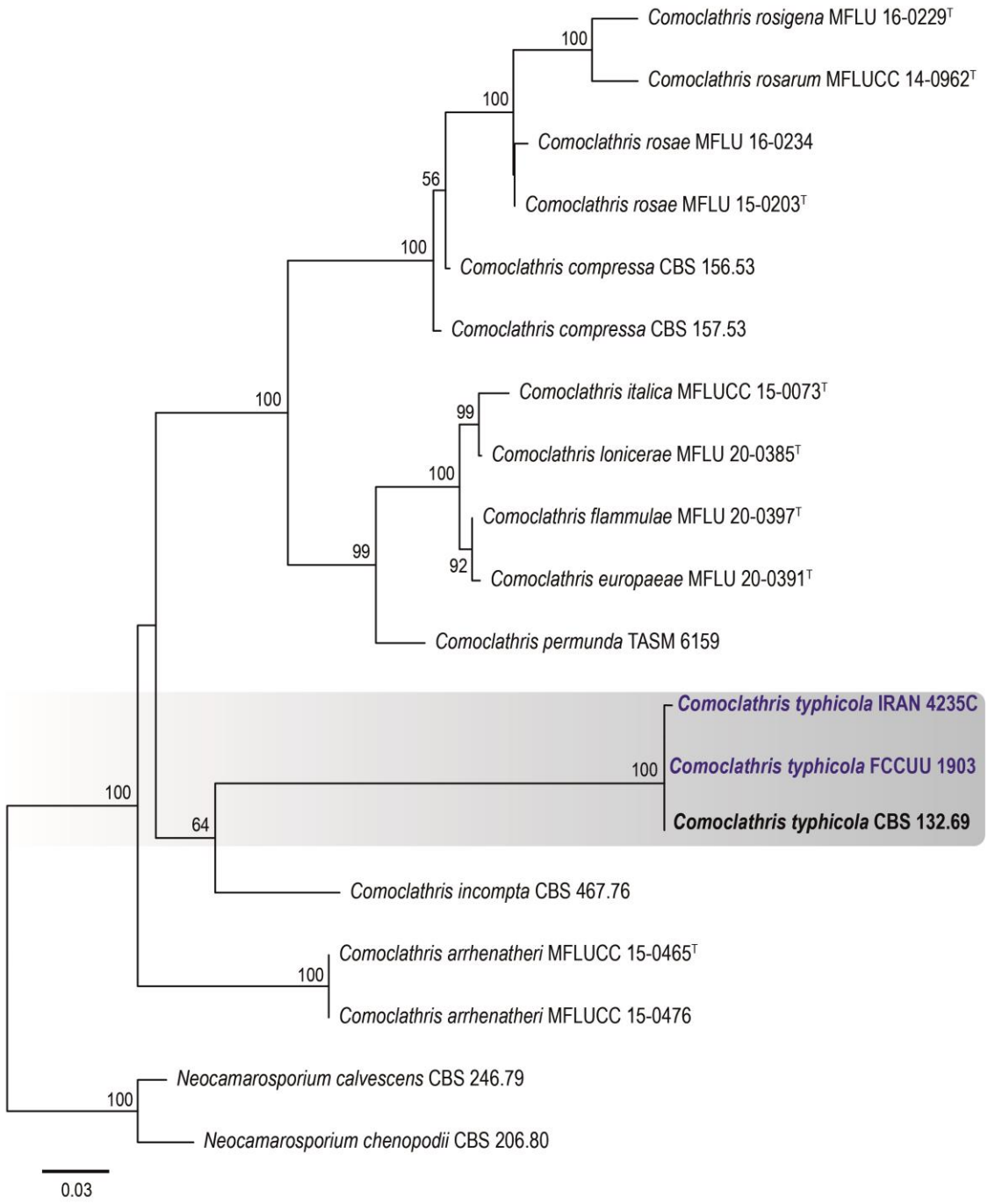

Fig. 1 Maximum Likelihood (ML) analysis based on *RPB2* sequence data of *Comoclathris* species. The Maximum Likelihood bootstrap support (BS) values >50% are given at the nodes. The tree was rooted to *Neocamarosporium chenopodii* (CBS 206.80) and *N. obiones* (CBS 432.77). The scale bar indicates the number of nucleotide substitutions. T indicates ex-type strains.

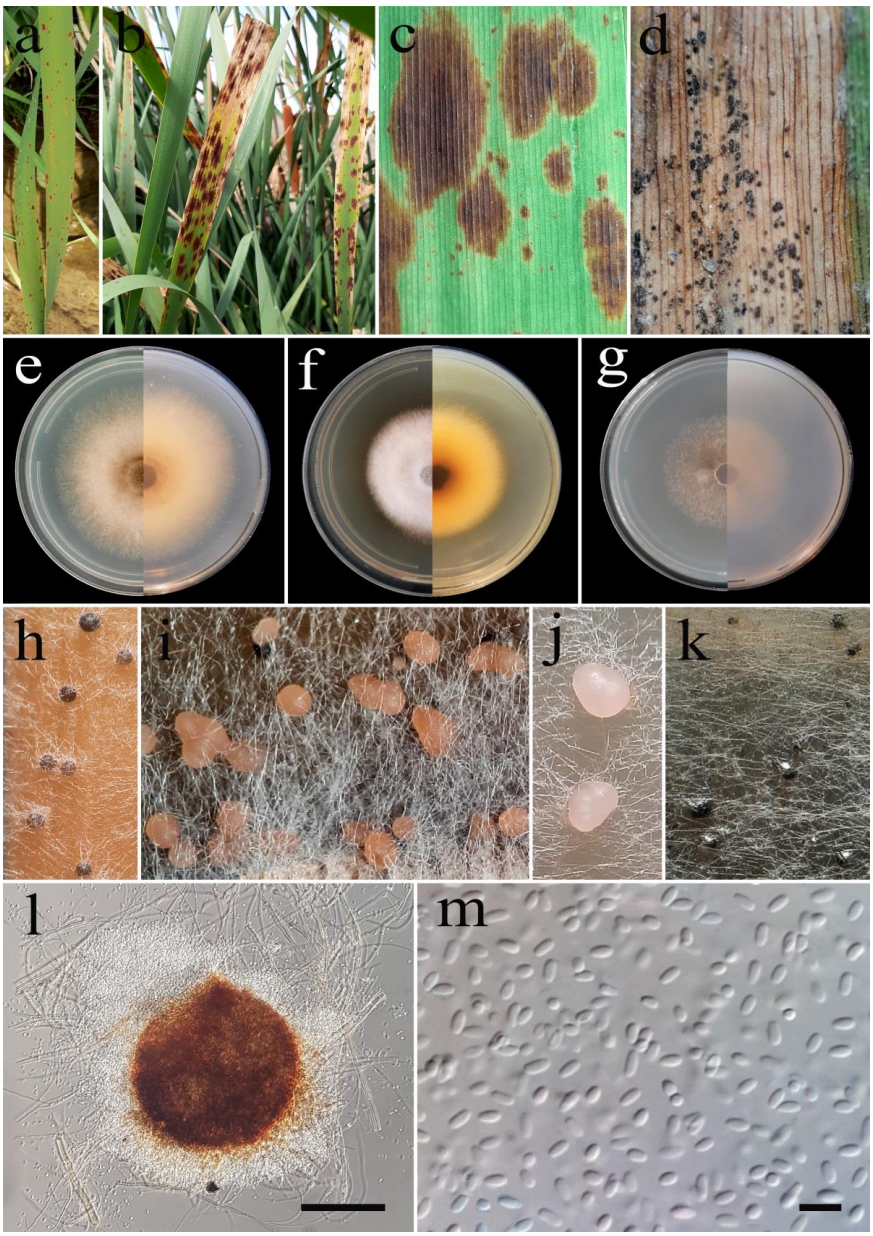

Fig. 2 *Comoclathris typhicola* (IRAN 4235C). **a–d.** Host plant showing necrotic lesions on leaves. **e-g.** Colonies (front and reverse) after seven days on PDA (e), MEA (f) and OA (g). **h–k.** Pycnidia with conidial mass on PDA (h-i) and OA (j-k). **l.** Pycnidium. **m.** Conidia. Scale bars: l = 100 µm, m = 10 µm.

Isolated from the leaves of *Typha latifolia* with necrotic lesions. Lesions up to 5–20 mm diam., spread on the upper surface, scattered, distinct, regular to irregular, pale brown to dark brown, leading to leaf death. Sexual morph: not observed. Asexual morph: Coelomycetous. Conidiomata pycnidial, semi-immersed to immersed, mostly solitary, rarely aggregated, scattered, globose to subglobose, pale brown to brown, thin-walled, glabrous, closed to one inconspicuous pore, with creamy to yellow conidial mass, 180–200 × 190–200 µm. Pycnidial wall pseudoparenchymatous, 3–5 layered, composed of oblong to isodiametric cells, pale brown, 10–25 µm thick. Conidiogenous cells phialidic, hyaline, smooth, globose to ampulliform, 4–6 × 4–7 µm. Conidia ellipsoidal to oblong, occasionally ovoid, with rounded ends, hyaline, smooth and thin-walled, aseptate, with polar guttules, 3–4 × 1–1.2 µm ($\bar{x}$ = 3.5 × 1.1 µm, n = 50).

Culture characteristics – Colonies on PDA reaching 57–59 mm diam. after seven days at 25 °C, smooth margin, with sparse aerial mycelia, grey at the center and white at the margin; reverse white to pale brown. Colonies on MEA reaching 49–50 mm diam. after

seven days at 25 °C, floccose, surface white, smooth margin, with sparse aerial mycelia; reverse white and pale brown at the center. Colonies on OA reaching 53–55 mm diam. after seven days at 25 °C, smooth margin, surface white to grey, with sparse aerial mycelia, abundant production of pycnidia, conidial matrix visible; reverse buff and grey near the center. Specimen examined – IRAN, West Azarbaijan province, Miyandoab city, isolated from infected leaves of *Typha latifolia* (Typhaceae, Poales), 10 Sept. 2021, *A. Ahmadpour*, IRAN 4235C (*RPB2* = OR611972) and FCCUU 1903 (*RPB2* = OR611973).

REFERENCES

Ahmadpour A, Ghosta Y, Poursafar A. 2021. Novel species of *Alternaria* section *Nimbya* from Iran as revealed by morphological and molecular data. Mycologia 113: 1073–1088.

Ariyawansa H, Phookamsak R, Tibpromma S, Kang JC, Hyde KD. 2014. A molecular and morphological reassessment of Diademaceae. The Scientific World Journal 2014: 1–11.

Ariyawansa HA, Thambugala KM, Manamgoda DS, Jayawardena R, Camporesi E, Boonmee S, Wanasinghe DN, Phookamsak R, Hongsanan S, Singtripop C, Chukeatirote E, Kang JC, Jones EBG, Hyde KD. 2015. Towards a natural classification and backbone tree for *Pleosporaceae*. Fungal Diversity 71: 85–139.

Boonmee S, Wanasinghe DN, Calabon MS, Huanraluek N, Chandrasiri SKU, et al. 2021. Fungal diversity notes 1387–1511: taxonomic and phylogenetic contributions on genera and species of fungal taxa. Fungal Diversity 111: 1–335.

Clements FE. 1909. The genera of fungi. Vol. 37. The HW Wilson Company. 227 pp.

Crous PW, Cowan DA, Maggs-Kolling G, Yilmaz N, Thangavel R, et al. 2021. Fungal planet description sheets: 1182–1283. Persoonia 46: 313–528.

Mattoo AJ, Ghosh A, Nonzom S. 2023. *Comoclathris acuminata* (Pleosporaceae, Pleosporales): A new endophytic species from Indian Himalayas. Phytotaxa 589: 230–244.

Miller MA, Pfeiffer W, Schwartz T. 2012. The CIPRES science gateway: enabling high-impact science for phylogenetics researchers with limited resources. Paper presented at: Proceedings of the 1st Conference of the Extreme Science and Engineering Discovery Environment: Bridging from the extreme to the campus and beyond (ACM).

Rambaut A. 2019. FigTree, a graphical viewer of phylogenetic trees. Available from: http://tree.bio.ed.ac.uk/software/figtree.

Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30: 1312–1313.

Wijayawardene NN, Hyde KD, Rajeshkumar KC, Hawksworth DL, Hugo M, et al. 2017. Notes for genera: Ascomycota. Fungal Diversity 86: 1–594.

Woudenberg JHC, Groenewald JZ, Binder M, Crous PW. 2013. *Alternaria* redefined. Studies in Mycology 75: 171–212.

Zhang Y, Koko TW, Hyde KD. 2011. Towards a monograph of Dothideomycetes: studies on Diademaceae. Cryptogamie Mycologie 32: 115–126.